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(71) Applicant: **GENE-TRAK SYSTEMS  
Two California Avenue  
Framingham Massachusetts(US)**

(72) Inventor: **Stackenbrandt, Erko  
Sturenhagener Weg. No. 5  
W-2307 Daenischenhagen(DE)  
Inventor: Curiale, Michael  
2751 Tarpon Court  
Homewood Illinois 60430(US)**

(74) Representative: **Deans, Michael John Percy et  
al  
Lloyd Wise, Tregear & CO. Norman House  
105-109 Strand  
London WC2R OAE(GB)**

(54) **Detection of listeria.**

(57) Nucleic acid fragments capable of hybridizing to  
rRNA of Listeria monocytogenes and not to rRNA of  
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**Framingham Massachusetts(US)**

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**Sturenhagener Weg. No. 5**  
**D-2307 Daenischenhagen(DE)**  
Inventor: **Curiale, Michael**  
**2751 Tarpon Court**  
**Homewood Illinois 60430(US)**

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## DETECTION OF LISTERIA

This invention relates to detecting bacteria belonging to the genus *Listeria*. (The term "*Listeria*," as used herein, refers to the bacteria classified as such in Bergey's Manual of Systematic Bacteriology, with the exception of *L. denitrificans*, which recently has been removed from the genus *Listeria* based on molecular and phenotypic consideration (Rocourt et al. (1987) International Journal of Systematic Bacteriology (submitted)). Detection of *Listeria* bacteria is important in various medical and public health contexts. *Listeria* infection can cause a variety of symptoms ranging from cold-like to flu-like, but is especially dangerous for a fetus, where it induces a 50% mortality rate.

According to a standard laboratory method and a method recommended by the F.D.A., the presence of *Listeria* in environmental or dairy specimens (e.g., milk) is detected by culturing an appropriately prepared sample on microbiological media under conditions favorable for growth of these organisms. The resulting colonies are examined for morphological and biochemical characteristics, a process that typically is started 48 hours after acquisition of the sample and takes between 9 - 19 days to complete.

Kohne et al. (1968) Biophysical Journal 8:1104-1118 discuss one method for preparing probes to rRNA sequences.

Pace and Campbell (1971) Journal of Bacteriology 107:543-547 discuss the homology of ribosomal ribonucleic acids from diverse bacterial species and a hybridization method for quantitating these homology levels.

Sogin, Sogin, and Woese (1972) Journal of Molecular Evolution 1:173-184 discuss the theoretical and practical aspects of using primary structural characterization of different ribosomal RNA molecules for evaluation phylogenetic relationships.

Fox, Pechman, and Woese (1977) International Journal of Systematic Bacteriology 27:44-57 discuss the comparative cataloging (of 16S ribosomal RNAs) approach to prokaryotic systematics.

The present invention will be better understood in light of the following definitions:

DNA - deoxyribonucleic acid, the type of nucleic acid containing deoxyribose as the sugar component.

RNA - ribonucleic acid, the type of nucleic acid containing ribose as the sugar component and which, most generally, is transcribed (copied) from DNA. RNA molecules may serve informational (e.g., messenger RNA), catalytic, or structural (e.g., ribosomal RNA, see below) cellular functions.

rRNA - Ribosomal RNA (rRNA) molecules are

key elements of ribosomes, complex protein and RNA-containing "organelles" which, together with transfer RNAs, comprise the translation apparatus. Ribosomes are of profound importance to all organisms because they serve as the only known means of translating genetic information into cellular proteins, the main structural and catalytic elements of life.

Ribosomes contain three distinct RNA molecules which, in *E. coli*, are referred to as 5S, 16S and 23S rRNAs. These names historically are related to the size of the RNA molecules, as determined by sedimentation rate. However, they actually vary substantially in size between organisms. 5S, 16S, and 23S rRNA are commonly used as generic names for the homologous RNA molecules in any bacteria, and will be so used here.

Hybridization - the process by which, under defined reaction conditions, two partially or completely complementary nucleic acids are allowed to come together in an antiparallel fashion and form specific and stable hydrogen bonds. The stringency of a particular set of hybridization conditions is defined by the base composition of the probe/target duplex, as well as by the level and geometry of mispairing between the two nucleic acids. Stringency may also be governed by such reaction parameters as the concentration and type of ionic species present in the hybridization solution, the types and concentrations of denaturing agents present, and/or the temperature of hybridization.

Probe(s) - synthetic or biologically produced nucleic acids (DNA or RNA) which, by design or selection, contain specific nucleotide sequences that allow them to hybridize, under defined stringencies, specifically (i.e., preferentially) to target nucleic acid sequences.

Target - a nucleic acid sequence to which a particular probe is capable of preferentially hybridizing.

Other definitions are given as their first use arises in the text.

The present invention features in one aspect nucleic acid probes or probe sets consisting essentially of DNA or RNA sequences which are capable, under specific hybridizing conditions, of detecting the presence of ribosomal RNA (rRNA) molecules of *Listeria monocytogenes* and are not capable, under the same conditions, of detecting the rRNA of closely related *B. subtilis* bacteria which may be present in the test sample.

The present invention also features in another aspect an assay system for the utilization of these probes, the format of which can enhance the afore-

mentioned desirable behaviour of the probes. Specific systems can produce the following enhanced capabilities:

- a) increased sensitivity; i.e., the ability to detect *Listeria* in a given sample more frequently than currently available methods,
- b) potentially significant cost reductions in probe production due to the use of chemically, rather than biologically, synthesized probes,
- c) accurate identification of even biochemically unusual *Listeria*, because of the rRNA sequence characterizations which provide the basis of such identification, and
- d) faster results because the test is performed on cultured cells which need not be grown further, the test of this invention taking only 2 - 4 days to provide a result.

The use of *Listeria* rRNA as target molecules provides a number of advantages, among them:

1) The rRNAs constitute a significant component of cellular mass. Although estimates of cellular ribosome content vary, actively growing *Listeria* bacteria may contain upwards of  $5.0 \times 10^4$  ribosomes per cell, and therefore  $5.0 \times 10^4$  copies of each of the rRNAs (present in a 1:1:1 stoichiometry in ribosomes). In contrast, most other potential cellular target molecules, genes or RNA transcripts thereof, are present in much lower abundance.

(2) The rRNAs (and the genes encoding them) appear not to be subject to lateral transfer between contemporary organisms. Thus, rRNA primary structure provides an organism-specific molecular target, rather than a gene-specific target as would likely be the case, for example, of a plasmid-borne gene or product thereof which may be subject to lateral transmission between contemporary organisms.

In addition to providing the advantages inherent in rRNA detection, the present invention provides probes to *Listeria* rRNA target sequences which are sufficiently similar in a significant number of *Listeria* that one or a few probes can hybridize to the target region in those *Listeria*, and are sufficiently different in most non-*Listeria* rRNAs that, under some conditions where the probe(s) hybridize to *Listeria* rRNAs, they are not capable of hybridizing, or hybridize very poorly, to most non-*Listeria* rRNAs. These probe characteristics are defined as inclusivity and exclusivity, respectively. In addition, the probes of the invention hybridize to target regions which can be rendered accessible to the probes under normal assay conditions.

In a particularly preferred embodiment of the invention, an assay method is used in which bacteria in the sample are grown under conditions which allow rapid growth of any *Listeria* in the

sample, but not of closely related *Brochothrix* species. Hybridization analysis is then performed on the sample after this growth period. In such an assay, some cross-hybridization of the probe with rRNA of the related *Brochothrix* bacteria can be tolerated because of the pre-hybridization selective amplification of the *Listeria*.

In another preferred embodiment, the *Listeria* detection method includes, prior to contacting the *Listeria* rRNA with one or more probes, the steps of subjecting the sample to a primary enrichment step for *Listeria* bacteria, followed by a secondary, non-specific bacterial enrichment step which is carried out in a buffer (most preferably, a 3- [N-Morpholino] propanesulfonic acid - containing buffer) to maintain the pH of the enriched sample between 6.5 and 8.0, most preferably about 7.2. This pH control, it has been found, permits the *Listeria* in the sample to grow much faster, during secondary enrichment, than is otherwise possible.

In another preferred embodiment, the *Listeria* detection method includes the step, prior to contacting the *Listeria* rRNA with one or more probes, of contacting the *Listeria* in the sample with an enzyme which weakens the *Listeria* bacterial cell walls, prior to subjecting the bacteria to lysis using a lytic agent, e.g., guanidinium thiocyanate. It has been found that this enzyme treatment greatly improves the hybridization reaction.

Other features and advantages will be apparent from the following description of the preferred embodiments.

Preferred embodiments of the invention are now described.

The Figure is the nucleotide sequence of five regions of *Listeria* 16S rRNA where *Listeria/B. subtilis* differences, shown, were discovered; corresponding rRNA sequences of *B. subtilis*, and other bacteria, are shown below *Listeria* sequences. L. mon represents *L. monocytogenes* (the *Listeria* species of greatest medical concern), L. inn represents *L. innocua*, L. mur represents *L. murrayi*, B.S. represents *Bacillus subtilis*, B.T. represents *Brochothrix thermosphactum*, P.V. represents *Proteus vulgaris*, E.C. represents *E. coli*, and H.V. represents *Halobacterium volcanii*.

#### Probe Development Strategy

We first identify regions of 16S and 23S rRNA which potentially can serve as target sites for *Listeria*-specific nucleic acid probes. As a practical matter, it is difficult to predict, a priori, which non-*Listeria* organisms might be present in any test sample. Because of the large number of such potential non-*Listeria* bacteria, demonstrating exclusivity for any given probe sequence is extremely

difficult and laborious. However, a more rigorous criterion can be adopted which obviates the need to know, during initial stages of research and development, what non-*Listeria* bacteria might be present in all test samples that ultimately will be screened using the probe. This entails a knowledge of the phylogenetic relationships among *Listeria* and between *Listeria* and other groups of bacteria. Specifically, it is taken as an operating hypothesis that the exclusivity criterion may be satisfied by determining that a particular target region in *Listeria* rRNA is sufficiently different from the homologous region in the rRNA of representative close evolutionary relatives of *Listeria* (in particular, *B. subtilis*) that the *Listeria* and relatives can be distinguished by hybridization using a probe specific to the *Listeria* sequence. As a general rule then, based on phylogenetic principles, the rRNA sequences of more distantly related organisms, even though their actual identity is not necessarily known, can be predicted to be at least as different, in a particular region of sequence, than the aforementioned close evolutionary relative of *Listeria*, *B. subtilis*.

As our first step in identifying regions of *Listeria* rRNA which might be useful as target sites for nucleic acid hybridization probes, we have determined nearly complete nucleotide sequences of the 16S rRNAs from three species of *Listeria*: *L. monocytogenes*, *L. innocua*, and *L. murrayi*. These were selected as representatives of the major *Listeria* DNA homology groups, and thus are representative of the evolutionary breadth of genus *Listeria*. Nucleotide sequences of various portions of the rRNAs were determined by standard laboratory protocols either by cloning (Maniatis et al. (1982) Molecular Cloning: A laboratory manual. New York: Cold Spring Harbor Laboratory, 545 pp.) and sequencing (Maxam et al. (1977) Proceedings of the National Academy of Science, USA 74:560-564; Sanger et al. (1977) Proceedings of the National Academy of Science, USA 74:5463-5467) the genes which specify the rRNAs, and/or by direct sequencing of the rRNAs themselves using reverse transcriptase (Lane et al. (1985) Proceedings of the National Academy of Science, USA 82:6955-6959).

These nucleotide sequences were compared to one another and to other available rRNA nucleotide sequences, in particular, to those of the closely related soil bacterium *Bacillus subtilis*. A number of regions were discovered that exhibit potentially useful exclusivity characteristics with respect to the *B. subtilis* rRNA sequences (i.e., contain *Listeria*-specific sequences). Certain of these are shown in the Figure, and discussed below.

As discussed above, this preliminary analysis provides only a demonstration of feasibility. Further experimental testing of each nucleic acid probe is

required to rigorously demonstrate the desirable characteristics discussed above, namely: 1) adequate exclusivity as to most or all closely related organisms, 2) useful inclusivity patterns with respect to *Listeria*, and 3) accessibility of the target regions under various assay conditions that might actually be employed. Because of the extremely large number of organisms potentially relevant to defining exclusivity and inclusivity characteristics of test probes, we have adopted the herein described interactive strategy with respect to testing and refinement of potential probes.

The first generation probes are designed based on the principle of maximizing utilization of the observed nucleotide sequence variation in the target regions of *Listeria* and non-*Listeria* bacteria. There is then carried out preliminary testing of the inclusivity and exclusivity properties of the first generation probes (which are synthesized by standard oligonucleotide techniques) by "dot blot" analysis. Dot blot analysis can be performed in many different ways, but generally involves immobilizing a nucleic acid or a population of nucleic acids on a filter (e.g., nitrocellulose, nylon, or other derivatized membranes which are commercially available and useful for this purpose). RNA can easily be so immobilized and then probed under any of a variety of nucleic acid hybridization conditions (i.e., stringencies), for nucleotide sequences of interest. Techniques are also available in which RNA present in crude (unpurified) cell lysates can be immobilized without having to first purify the nucleic acid in question. This latter approach significantly decreases the amount of effort required to screen for particular nucleotide sequences which may be present in the nucleic acids of any particular organism and, moreover, is amenable to the mass screening of large numbers of organisms. It, therefore, is the method of choice for testing the exclusivity and inclusivity properties of potential nucleic acid hybridization probes versus large numbers of organisms.

A list of non-*Listeria* bacteria which exemplify the type of bacteria that may be present in potentially *Listeria*-containing samples is given in Table 1. These also represent many of the genera most closely related to *Listeria*. As discussed above, a probe which demonstrates good exclusivity characteristics to such a broad representation of bacteria can reasonably be predicted to behave similarly to a much broader list of organisms than actually tested. DNA probes 568, 609, and 661 (Figure) did not hybridize to over 90% of these bacteria, under the stringency conditions described in the Example, below. (Probes 568, 609, and 661, and the other probes of the Figure, are complementary (5' probe end to 3' RNA end) to the given *L. monocytogenes* rRNA sequences, i.e., the

probes each have a C for the RNA's G, a T for the RNA's A, an A for the RNA's U, and a G for the RNA's C.)

Table 1

Partial Listing of Non-Listeria Bacteria Used in Screening Potential Probes

Aeromonas sobria  
Bacillus cereus  
Bacillus subtilis  
Brochothrix thermosphacta  
Citrobacter freundii  
Corynebacterium xerosis  
Corynebacterium diptheriae  
Escherichia vulneris  
Enterobacter agglomerans  
Enterobacter cloacae  
Klebsiella pneumoniae  
Klebsiella oxytoca  
Lactobacillus casei  
Pseudomonas aeruginosa  
Rhodococcus equi  
Salmonella arizonae  
Salmonella cholerae-suis  
Salmonella typhi  
Serratia odorifera  
Shigella boydii  
Shigella flexneri  
Shigella sonnei  
Staphylococcus aureus  
Staphylococcus epidermidis  
Staphylococcus hominis  
Staphylococcus saprophyticus  
Streptococcus agalacticae  
Streptococcus bovis  
Streptococcus faecalis  
Streptococcus faecium  
Streptococcus lactis  
Streptococcus mutans  
Streptococcus pneumoniae  
Streptomyces globisporus  
Yersinia enterocolitica

Probes 568, 609, and 661 (Figure) also exhibited good inclusivity properties, by binding, under the stringency conditions of the Example, to *L. monocytogenes*, *L. innocua*, *L. murrayi*, *L. ivanovii*, *L. seeligeri*, *L. welshimeri*, and *L. grayi*.

Several other considerations also affect optimal design characteristics of a probe sequence. The first is consideration of the geometry of the probe with respect to itself (i.e., intramolecular interac-

tions). Potentially useful target regions of the 16S rRNA of *L. monocytogenes* may be located in regions that exhibit substantial potential for self complementarity. Therefore, probes to these regions can also exhibit self complementarity. Because potential interactions between the probe and target sequences are governed by the same types of interactions that govern the intramolecular annealing of the target or probe sequences to themselves, it is possible, particularly under solution hybridization conditions, that self-complementary probes can render themselves inaccessible for hybridization to their target sequences. Thus, one aspect of probe design is to minimize such self-complementarity. This can necessitate making a compromise between maximum utilization of *Listeria*-specific sequences and acceptable probe geometry.

An additional consideration in probe design arises with respect to the inclusivity criterion. Preferred probes are those which, while displaying appropriate exclusivity behavior, also can hybridize to the rRNA(s) of all desired *Listeria* bacteria. Because the genus *Listeria* itself is comprised of bacteria which exhibit significant phenotypic and genetic variations, it may not be possible to design (or discover) such an "ideal" probe. In practice, rather than demanding a "universal" *Listeria* probe, a set of *Listeria*-specific probes is sought, each of which exhibits appropriate exclusivity, and some useful level of inclusivity. In aggregate, the set of probes will detect most or all *Listeria* and few or no non-*Listeria* bacteria. In such a set, for example, one probe can detect all but one or a few important *Listeria* strains, and another probe may hybridize with good exclusivity only to those few *Listeria* strains missed by the first probe. Thus, although the probes disclosed below are characterized on an individual basis with respect to inclusivity characteristics, it must be borne in mind that the concept of "sets" of specific probes detailed above must also be considered in determining the importance of individual probes.

This set notion also extends to evaluation of the exclusivity behavior of individual probes. For example, in the detection format described below, undesirable hybridization to non-*Listeria* bacteria by otherwise useful probes or probe sets can be reduced or abolished by manipulation of the assay strategy employed.

In some cases, it is preferred to have probes specific for just one species of *Listeria*, specifically *L. monocytogenes*, which is pathogenic to animals and especially to humans. Such probes are useful since they will not produce false positive results when non-pathogenic *Listeria* are in a sample, and thus a positive result indicates the presence of pathogenic *Listeria*.

The final step of probe design and analysis is testing of real (e.g., food/clinical) samples.

#### Probes

The probe selection strategy described above has yielded a number of probes useful in identifying *Listeria* bacteria in samples. The first step in the probe selection process was to carry out nucleotide sequence analysis on the 16S rRNA of the three *Listeria* spp. listed above. Comparison of these *Listeria* sequences to non-*Listeria* rRNA sequences (particularly *B. subtilis* sequences) identified sequences which meet the exclusivity criterion, i.e., they are essentially *Listeria* specific, and are thus potential target regions to which probes could be directed. These regions are indicated in the Figure.

Liquid hybridization assays using probes 568 and 661 (the sequences of these probes are 568: 5' TCGCGGCTTCGCGACCTTTGTACTATCCA 3' and 661: 5' GGGAAAGCTCTGTCTCCAGAGTGGTCAAAGG 3') demonstrate good accessibility of target sequences in these regions under different hybridization conditions. Generally, good sensitivity also is achieved. However, these probes will hybridize under the above conditions to *Brochothrix*, and thus these bacteria must be selected against during the assay procedure. One method is to culture bacteria within a sample under conditions which do not allow *Brochothrix* to grow, e.g., at temperatures above 30° C, such as 32° C or 35° C. In this way, false positive results are avoided because *Brochothrix* background is very low.

Probe 609 does not hybridize under the above conditions to *Brochothrix* and thus selective *Listeria* amplification is not needed where this probe is used as the only probe.

Probes specific to *Listeria* 23S rRNA can be derived as described above in the case of 16S rRNA.

The probes of the invention can be used in a variety of formats, one of which is described below.

#### Example

The DNA hybridization test employs *Listeria*-specific DNA probes labelled with a radioisotope (phosphorus 32, although other labels are equally suitable, e.g., fluorescent or chemiluminescent labels) to detect *Listeria* spp. in meat, dairy products and environmental samples following broth culture enrichment. After enrichment, bacteria present in the test samples are collected on membrane filters by vacuum filtration. The bacteria are lysed and the released nucleic acids are fixed to the membrane

filters. Following a short pre-hybridization step, the filters are incubated in a hybridization solution to which the <sup>32</sup>P-labelled *Listeria*-specific DNA probes are added. If *Listeria* nucleic acids (ribosomal RNA) are present in a test sample, the radiolabelled DNA probes will hybridize to the target nucleic acid sequences present on the filter. Unbound probe is washed away and the radioactivity on the filters is measured by counting in a scintillation counter or other beta particle detector. Radioactivity on a filter above a threshold value, determined by results obtained for negative control assays, indicates the presence of *Listeria* in the test sample. The assay is a qualitative test to determine the presence of *Listeria* spp. in dairy or meat products and environmental samples. A sample is considered non-reactive for the presence of *Listeria* spp. if the counts per minute (cpm) value does not exceed the established cutoff value for the assay. A sample is considered reactive for the presence of *Listeria* spp. if the cpm value is greater than the established cutoff value for the assay.

The first step was primary enrichment for *Listeria* in the sample. For dairy product samples, 25 g of food sample was added to 225 ml of *Listeria* Enrichment Broth (EB; Trypticase soy broth powder 30 g, Yeast extract 6 g, Distilled water 1 liter, Acriflavin HCl 15 mg/liter, Nalidixic acid (Na salt) 40 mg/liter, and Cycloheximide 50 mg/liter.) The mixture was homogenized using a blender or stomacher as appropriate for the particular sample type, and then incubated in a flask or bottle for 24 ± 4 hours at between 30° C and 50° C, preferably 35° C.

For environmental testing, a swab or other environmental sample was placed in a flask or bottle containing 25 ml (or larger volume if required) EB, and incubated for 24 ± 4 hours at between 30° - 50° C, preferably 35° C.

The next step was secondary enrichment for all sample types. The primary enrichment culture was removed from incubation and mixed well. 1 ml of the primary enrichment culture was transferred to a flask or bottle containing 100 ml modified *Listeria* Enrichment Broth (MEB; Trypticase soy broth powder 30 g, Yeast extract 6 g, 3-[N-Morpholino] propanesulfonic acid -free acid (MOPS -free acid) 8.5 g, 3-[N-Morpholino] propanesulfonic acid - Na salt 8.5 g, MOPS - Na salt 13.7 g, Distilled water 1 liter, Acriflavin HCl 15 mg/liter, Nalidixic acid (Na salt) 40 mg/liter, and Cycloheximide 50 mg/liter) and incubated for 24 ± 4 hours at 30° - 50° C, preferably 35° C.

The following assay procedure makes use of the GENE-TRAK manifold kit (GENE-TRAK Systems Cat. No. GT801). One ml of the cultured cells were treated with 0.5 ml of enzyme solution (containing 3000 units of mutanolysin (available



from Sigma) and 150 mg of lysozyme (available from Sigma) in 15 ml of 10xTE (Tris 10mM, EDTA, 1mM, pH 7.6) buffer by incubation at room temperature (15 - 30° C) for 15 min. The contents of each tube were then vacuum filtered into a filter cup. This results in a concentration on the filter of the cultured cells, the cell walls of which have been weakened by the enzymes.

The nucleic acid in these cells was then denatured as follows. *Listeria* lysing and denaturing solution (0.675 M guanidine thiocyanate, 1xTE) was squirted into the filter cups until the filter was covered with liquid and after 2 minutes the solution vacuum filtered. *Listeria* neutralizing solution (1 M Tris pH 8, 0.6 M NaCl) was then squirted into the filter cup until the filter was covered with liquid, and again after 2 minutes the solution was vacuum filtered. A *Listeria* fixation solution (95% ethanol, 5% water) was then squirted into the filter cup until the filter was covered with liquid and after 2 minutes the solution vacuum filtered.

The denatured nucleic acid on the filter was then hybridized, as follows. The filters were removed from the cups and placed in a 50 ml conical bottom tube containing 25 ml of *Listeria* prehybridization solution (6xSSC, 0.5% SDS, 1xDenhardtts, 1mM EDTA) prewarmed to 65° C. The tube and contents were placed in a 65° C water bath for 30 minutes. The liquid was poured out of the tube and 12 ml of *Listeria* hybridization solution (6xSSC, 0.5% SDS, 1x Denhardtts, 1mM EDTA and 12  $\mu$ Ci of labeled *Listeria* probe) prewarmed to 65° C added. The tube and contents were placed in a 65° C water bath for 120 minutes.

After hybridization, the filters were washed as follows. The contents of the tube were discarded and 25 ml of *Listeria* wash solution (0.5xSSC, 0.1% SDS, 0.16% antifoam B), prewarmed to 65° C, was added. The tube was shaken and placed at 65° C for 5 minutes. The tube was then again shaken and the liquid poured out. This process was repeated five more times.

The washed filters were removed from the tube and placed on a clean sheet of paper to count in a beta-counter for 30 seconds, and the cpm for each filter recorded.

Generally the average of 3 negative control filters (prepared from a selected non-*Listeria* bacterium) was taken and 500 cpm added to this value. This number is defined as the cutoff value. Filters with more cpm are suspect for *Listeria*, those with less indicate the absence of *Listeria*.

## Claims

1. A nucleic acid fragment capable of hybridizing, under predetermined stringency conditions, to rRNA of *Listeria monocytogenes* and not to rRNA of *Bacillus subtilis*.

2. The nucleic acid fragment of claim 1 wherein said fragment is not capable of hybridizing, under said conditions, to rRNA of *Aeromonas sobria*, *Bacillus cereus*, *Bacillus subtilis*, *Brochothrix thermosphacta*, *Citrobacter freundii*, *Corynebacterium xerosis*, *Corynebacterium diptheriae*, *Escherichia vulneris*, *Enterobacter agglomerans*, *Enterobacter cloacae*, *Klebsiella pneumoniae*, *Klebsilla oxytoca*, *Lactobacillus casei*, *Pseudomonas aeruginosa*, *Rhodococcus equi*, *Salmonella arizonae*, *Salmonella cholerae-suis*, *Salmonella typhi*, *Serratia odorifera*, *Shigella boydii*, *Shigella flexneri*, *Shigella sonnei*, *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Staphylococcus hominis*, *Staphylococcus saprophyticus*, *Streptococcus agalactiae*, *Streptococcus bovis*, *Streptococcus faecalis*, *Streptococcus faecium*, *Streptococcus lactis*, *Streptococcus mutans*, *Streptococcus pneumoniae*, *Streptomyces globisporus*, *Yersinia enterocolitica*.

3. A set of nucleic acid fragments comprising a first nucleic acid fragment capable of hybridizing to rRNA of a first *Listeria* species and not to rRNA of *B. subtilis* or to rRNA of a second *Listeria* species, and

a second nucleic acid fragment capable of hybridizing to rRNA of said second *Listeria* species and not to rRNA of *B. subtilis*.

4. The set of nucleic acid fragments of claim 3, said fragments together being capable of hybridizing to at least 80% of *Listeria* strains.

5. The nucleic acid fragment of claim 1, said fragments being capable of hybridizing under hybridizing conditions to probe 568 or its complementary strand.

6. The nucleic acid fragment of claim 1, said fragments being capable of hybridizing under hybridizing conditions to probe 661 or its complementary strand.

7. The nucleic acid fragment of claim 1, said fragment being homologous with at least 75% of a 10 nucleotide or larger region of one of the five *Listeria* 16S rRNA regions shown in the Figure.

8. A method of detecting the presence of *Listeria* bacteria in a sample, comprising contacting the nucleic acid fragment of claim 1 with bacteria in said sample under conditions that allow said fragment to hybridize to rRNA of said *Listeria* bacteria in said sample to form hybrid nucleic acid complexes, and



detecting said hybrid complexes as an indication of the presence of said Listeria bacteria in said sample.

9. The method of claim 8 comprising culturing bacteria in said sample under culture conditions which allow growth of Listeria but not of Brochothrix.

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10. The method of claim 9 wherein said culture conditions is a growth temperature above 30° C.

11. A method for detecting Listeria in a sample comprising bacteria, said method comprising growing said bacteria at a temperature above 30° C, and then contacting, under hybridizing conditions, the rRNA of said bacteria with a nucleic acid probe able to hybridize to rRNA of L. monocytogenes and rRNA of Brochothrix but not to rRNA of B. subtilis, and detecting hybrid complexes as an indication of the presence of Listeria in said sample.

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12. The method of claim 8 wherein, prior to said contacting, said sample is subjected to a primary enrichment step for Listeria, followed by a secondary, non-specific bacterial enrichment step, said secondary enrichment step being carried out in a buffer solution to maintain the pH of said enriched sample between 6.5 and 8.0.

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13. The method of claim 12 wherein said pH is maintained at about 7.2

14. The method of claim 8 wherein, prior to said contacting, Listeria in said sample are contacted with an enzyme to weaken their cell walls, and are then lysed.

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15. A set of nucleic acid fragments derived from Listeria and having nucleotide sequences as shown in the accompanying drawings.

16. A nucleic acid fragment derived from Listeria and having any of the nucleotide sequences shown in the accompanying drawings.

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Nouvel dépôt / Newly filed  
nouveau dépôt

```

:-----IG 563-----:
L. monocytogenes  AAGCGCGCGGAGGCGGUCUUUUA
B. subtilis       ---G-U-----U-C---
Brochothrix       -----C-----
Proteus vulgaris  -----A-----AA---
E. coli           -----A-----U-G---
Halobacterium     -----UC--U--C--C-ACGA-G

```

```

:-----IG 565-----:
L. mon  UCGGAUUGUAGGCUGCAACUCGCCUGCAUGAAGCCGGAA
B.S.    -----C-C-U-----A---G-----U---
B.T.    -----C-----N-----
P.V.    C-----G-U-----A-C-----U---
E.C.    C-----G-U-----A-C-----U---
H.V.    -----AG-----A-----CU-----U--U

```

```

:-----662-----:
:-----661-----:
:-----564-----:-----566-----:
L. mon  UUACAUCUUUGACCACUCUGGAGACAGAGCUUUCUUUCGGGGACAAAGU
L. inn
L. mur
B.S.    ---C---A-UC-A---U-GA-G-C-----G-G---
B.T.    -----U-----A-G-----
P.V.    --AGC--AUC--U-A---U---GAG-G-----A-GCU-A
E.C.    --ACG--AGUU-UCA---UGAGAA-C G-----A--CGU-A
H.V.    A-AG---UG--GA-C --GUU-- --GA--U-AUCAC---GCU--

```

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:-----663 complement 568-----:
:-----610-----:
:-----568-----:-----567-----:
L. mon  UGGAUAGUACAAAGGGUCGCGAAGCCGCGAGGUGGAGCUAA UCCCAUAAAACUAUUCUCAG
L. inn  ---G-----N---C---C-----
L. mur  ---GA-----N---N---U-----
B.S.    ---C-A-----CA---A-----UA-C-U-----C-U-G---
B.T.    ---A-----U-----N---C-----U-----
P.V.    --CAGA-----A-AA---CCU-----AGCA--GG- A-U-----GUCUG--GU--
E.C.    --CGCA-----A-AA---CCU-----AGCA--GG- C-U-----GUGCG--GU--
H.V.    --UCGAG---U---U-U-UCU-AA-AGAAC----- --U-C---CUCGA--GU--

```

```

:-----609-----:
L. mon  CCGAAGUCGGUAGGGUACCUUUAUGGAGCCAGCCGC
L. inn  -----N-----
L. mur  -----A-----U---
B.S.    -----GA---A---UA-----
B.T.    --A---C---UU---A---CG---U---U
P.V.    AA-----A---CU-A-----
E.C.    AA-----A---CU-A-----

```